

# Manual for *hierarchicalBoosting* package

Marc Pybus, Pierre Luisi, Giovanni Dall’Olio, Manu Uzkudun,  
Hafid Laayouni, Jaume Bertranpetit and Johannes Engelken \*

May 5, 2015

This manual explains how to use the ‘Hierarchical Boosting’ method described in the manuscript entitled “A Machine-Learning Framework to Detect and Classify Hard Selective Sweeps in Human Populations”. Check <http://hsb.upf.edu/> for more information on citation.

## 1 Principle

The basic idea of the framework provided in this package is described in detail in the manuscript mentioned above, please read it before using it. The manuscript describes a framework to analyze and classify positive selection in genomic regions using polymorphism data. The novelty of the approach lies in the way the selection analysis is performed. Instead of addressing the inference of positive selection as a binary outcome (selection vs. non-selection), it assumes that there are different selective scenarios for which different selection tests show different power to detect. Under this assumption, it is possible to use this disparity in sensitivity towards different selection regimes to classify hypothetical selective sweeps into further properties, such as sweep age, completeness or intensity.

Our package is based on a machine-learning classification algorithm called *boosting* (from the *mboost* package - Buehmann and Hothorn 2007). *Boosting* is a supervised algorithm that estimates linear regressions (we call it *boosting* functions) of input variables (summary statistics of selection tests) to maximize the differences between two competing scenarios (e.g. complete vs. incomplete selective sweeps). Our method sequentially applies different boosting functions (as used by Lin *et al.* 2011) into a hierarchical classification scheme to optimally classify genomic regions into different selection regimes. The framework implemented here relies on the results of several selection test that need to be previously computed on simulated and empirical data. We do not provide the software needed to estimate selection tests or run the simulations.

New methods to detect selection are published every year improving the capabilities of previous ones or expanding the range of selection scenarios detected (hard and soft sweeps or balancing selection). We advise to use as many different selection tests as possible in the algorithm training process. High correlation (e.g.  $r^2 > 0.8$ ) between summary statistics or selection tests should be avoided since it will prevent coefficient convergence of the implemented boosting algorithm (as explained in the manuscript).

During the training process, the algorithm is fed with a table containing selection tests scores or summary statistics (as columns) for the different selection scenarios to be classified (as rows). The more replicates are used in the training process, the better. Table 1 shows an example of such input table. Note that an even number of replicates for the two competing scenarios is not required as explained in the manuscript.

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\*Institut de Biologia Evolutiva (UPF-CSIC), Universitat Pompeu Fabra, Barcelona, Spain. Contact:

scenario	CLR	iHS	FayWuH	TajimasD	FuLiD	XPEHH
neutral	0.2789	0.4424	-15.6810	-0.5916	0.6760	0.6449
neutral	0.7685	0.5439	2.2182	-0.2074	0.6502	-0.7827
...	...	...	...	...	...	...
ancient_complete	3.3130	1.1149	0.4764	2.23085	-3.5784	3.1230
ancient_complete	11.2205	1.6295	1.3220	-2.4338	-3.8018	4.0536
...	...	...	...	...	...	...
recent_incomplete	0.4207	1.8648	-21.6999	-1.7474	-0.44682	6.0771
recent_incomplete	0.5756	1.3418	-24.5180	-1.5016	-0.05735	8.3427

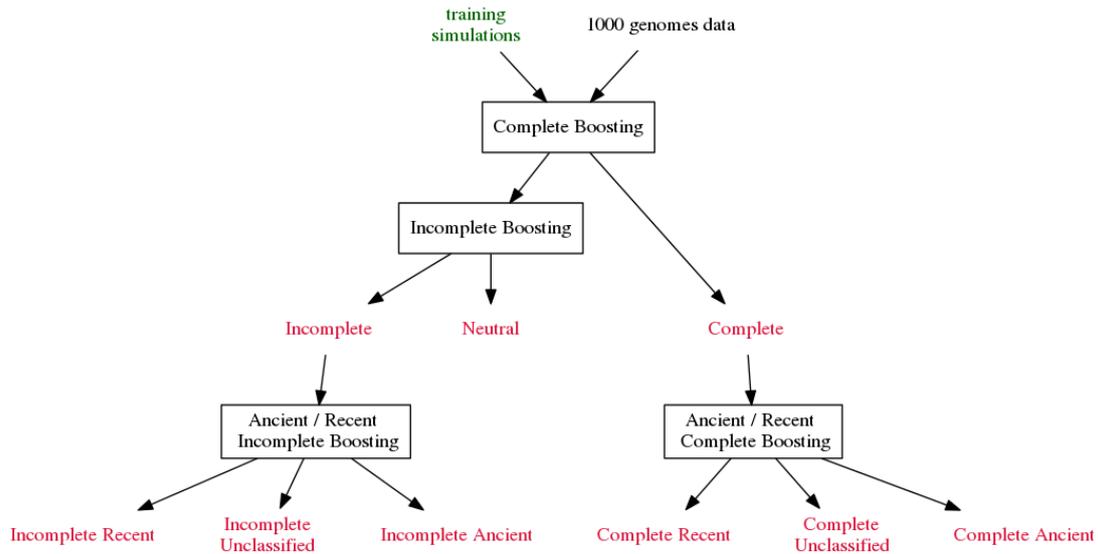
**Table 1.** Training table example.

## 2 Usage

Several steps need to be followed to properly use this package. This is a brief description of the typical selection detection and classification procedure:

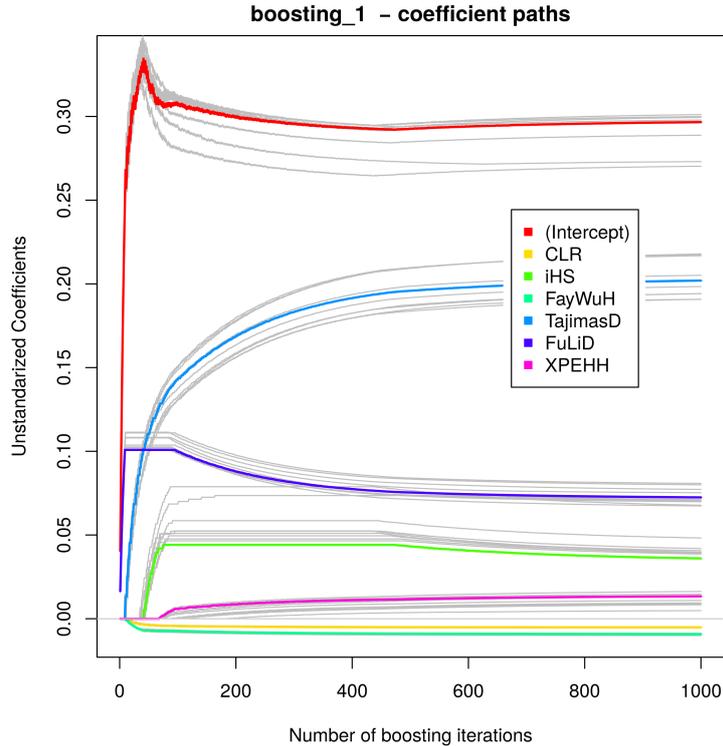
1. *Decide which selection tests to use:* depending on the type and properties of the data to analyze, different selection test should be used. Some tests allow the use of missing data, others need complete genotyping data. Some test may show sensitivity to many types of selection while others only to very specific ones. You should think which type of selection you are trying to detect (or discard) and choose your selection tests accordingly.
2. *Run simulations for the training process:* simulations should be as similar as possible to the empirical dataset you want to analyze. You should use state-of-the-art simulation software allowing for complex demographic and genomic models. Simulators should recreate different selective regimes (positive, negative, balancing selection) at different time-frames and populations. We recommend using one of the following simulation packages: SFS\_CODE (Hernandez 2008), SLiM (Messer 2013), msms (Ewing and Hermisson 2010) or cosi (Schaffner *et al.* 2005). Simulated data should mimic real genomic regions and match sample sizes and possible site frequency spectrum bias of your reference dataset. Specific recombination maps for the analyzed genomic regions should also be considered.
3. *Apply your selection tests to the simulated and empirical data:* selection tests usually come in different flavors: some are SNP-based, others are window-based, some contain several internal parameters that need to be tuned according to your data, others are applied as provided. Nonetheless, at the end you want to obtain a unique score per test and per region so they can be properly combined in a linear regression. We advice to use some sort of summary statistics (mean, maximum, minimum) to unify your input variables in a given genomic region (empirical or simulated). In the manuscript we describe an example of strategy to choose the most sensitive summary statistics.
4. *Define the classification tree in which data will be sequentially classified:* you need to envision which classification structure will provide the best results. You can also try several configurations and pick the best performing one. One good strategy is to first distinguish neutral from selection scenarios, and in the following steps, classify the selection scenarios according to common features, such as age, sweep completeness or intensity. The power of your selection tests towards specific selective scenario should guide you on the structure of

the classification tree. Remember that each node is a specifically trained *boosting* algorithm that maximizes different between two competing scenarios (Figure 1). While developing the method we realized that final allele frequency (Faf) of the allele under selection was driving the signal in almost all the selection tests implemented, as explained in the manuscript. Thus, we decided to first distinguish complete selective sweeps (Faf=1.0) compared to incomplete and neutral scenarios, which were classified afterward. This strategy proved to be better than discarding neutrality as the first step.



**Figure 1.** *hierarchicalBoosting* classification tree structure (from the manuscript).

5. *Generate validation plots and interpret them:* we have implemented a function that generates three validation plots for each trained *boosting* function. First validation plot is coefficient convergence plot (Figure 2). Here you must check that coefficients for your input variables reach convergence within the *boosting* iterative process and that the estimated coefficients are similar across bootstraps. Lack of convergence may appear due to high correlation between input variables or low number of *boosting* iterations. Correct this when convergence is not reached with the default parameters. A second validation plot is standardized coefficient plot (Figure 3). This plot shows the standardized coefficients for each *boosting* function, which can give you an insight about the importance of each selection test to uncover a specific selection scenario. The amplitude of the boxplot reflects the robustness of the estimated *boosting* function (generated by the bootstrapping process). To reduce variance in the coefficient estimation you can increase the number of replicates for each scenario or increase the number of bootstraps. And finally, the third validation plot informs about the distribution of your *boosting* scores in scenario A and scenario B, along with the significance threshold used (Figure 4). In this plot one can check how good is the estimated *boosting* function in distinguishing between the two competing scenarios, as well as which significance thresholds are more appropriate for a given *boosting* function.
6. *Evaluate performance of the trained hierarchicalBoosting tree:* once you have your *boosting* functions trained, validated and embedded in the classification tree (we call it a *hierarchicalBoosting*



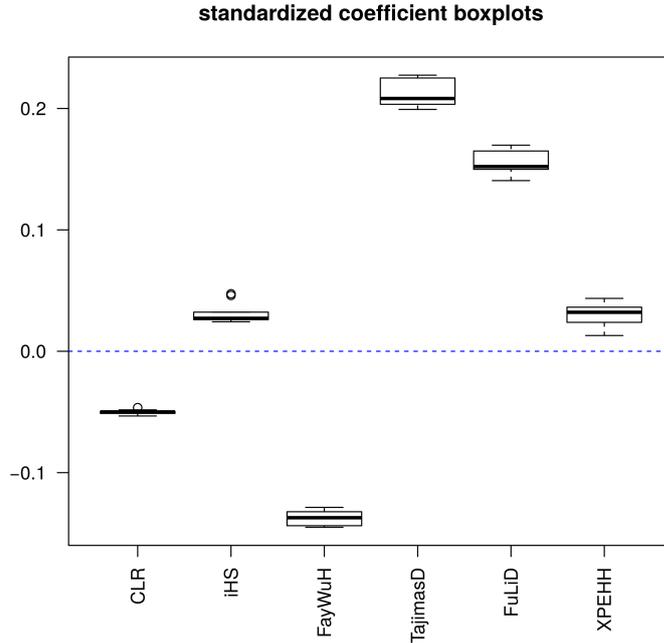
**Figure 4.** Coefficient convergence plot for the *boosting* function #1 from the example.

object), you should evaluate it before applying it to real data. Generate several independent evaluation simulations (with the scenarios to be classified or possible false positives) and apply the trained *hierarchicalBoosting* object to evaluate its classification power.

7. *Apply the trained hierarchicalBoosting tree to your empirical data:* the final step is to scan real data to detect and classify selection signals using your trained *hierarchicalBoosting* object. The simulations used to train the method should be as similar as possible to the empirical dataset. Any know bias should be specifically simulated.

### 3 Example

A full example is shown below. In this example the training data comes from the original manuscript (although it is not the exact same dataset). The data consists in 5 different selection scenarios generated using the *cosi* package (Schaffner *et al.* 2005). For each scenario 100 replicates are used. The simulated data mimics some population genetics features shown by the CEU population from The 1000 Genomes Project - Phase I dataset (The 1000 Genomes Project Consortium 2012). The selection scenarios simulated are: neutral, recent incomplete selection, recent complete selection, ancient incomplete selection and ancient complete selection. *Complete* and *incomplete* scenarios refer to the frequency of the allele under selection at the time of sampling; *incomplete* refers to selective sweeps with a final allele frequency of 0.8 ( $s = 0.0147$ ),



**Figure 4.** Standardized coefficient plot for the *boosting* function #1 from the example.

and *complete* specify selective sweeps that already reached fixation ( $s = 0.0257$ ). *Recent* and *ancient* scenarios indicate when those selective sweeps happened, being *recent* a sweep ending at the moment of sampling, and *ancient* a sweep that ended 30,000 years ago (generation time = 25 years).

The selection tests used here are the following: CLR (Nielsen *et al.* 2005), iHS (Voight *et al.* 2006), Fay & Wu’s *H* (Fay and Wu 2000), Tajima’s *D* (Tajima 1989), Fu & Li’s *D* (Fu and Li 1993), and XPEHH (Sabeti *et al.* 2002). Those tests were run under test-specific internal parameters (window size, MAF filtering,...) and then summarized in a common window size (using minimum or maximum, or mean) as described in detail in the manuscript. A window size of 25 Kbp was used to unify the selection tests.

The obtained data (selection tests scores for the simulated selection scenarios) was merged into a table that can be found in "exdata" directory in the package. This table is used as training dataset for the *train.hierarchicalBoosting* function.

A similar table containing selection tests scores for a real genomic region around the gene SLC45A2 (known to be under selection in CEU population) can be found in the same folder. We use this dataset as an example of empirical genomic region to be analyzed with the *apply.hierarchicalBoosting* function once the training step is finished. This dataset was generated by applying the same selection tests and the same summarizing approach used to create the training dataset.

A configuration table containing the parameters to apply the *hierarchicalBoosting* algorithm must be created (Table 2). It describes the relation between the inputs and outputs of each *boosting* function (Figure 1), the competing scenarios used, and the significance thresholds needed

	input_data	scenario_A	scenario_B	threshold_A	threshold_B
boosting_1	original	recent_complete, ancient_complete	neutral,recent_incomplete, ancient_incomplete	FALSE	0.01
boosting_2	boosting_1:scenario_B	neutral	recent_incomplete, ancient_incomplete	0.99	FALSE
boosting_3	boosting_1:scenario_A	recent_complete	ancient_complete	0.95	0.05
boosting_4	boosting_2:scenario_B	recent_incomplete	ancient_incomplete	0.95	0.05

**Table 2.** Configuration table for *hierarchicalBoosting*.

to classify the different selection scenarios. Thus, to run a given *boosting* function one must define the following parameters:

1. *input\_data*: the first *boosting* function must use as input dataset the original training table. This way, the following *boosting* functions can use the outputs from boosting #1 (referred as scenario A and scenario B) to further classify the data. When two thresholds are given to a *boosting* function a third classification outcome can be produced (scenario C). Figure 2 shows the score distribution of a typical *boosting* function in two competing scenarios with two significance thresholds used.
2. *scenario\_A* and *scenario\_B*: here one must describe the competing scenarios to be used at each *boosting* function. Scenarios names should be present in the training dataset and be comma-separated (as shown in Table 2).
3. *significance thresholds for each scenario*: values for the significance thresholds (between 0 and 1) for scenario A and scenario B. This allows to classify empirical data into the two or three possible outcomes. When only one threshold is used (to discard neutrality, for example), the other one must be set as FALSE.

Finally, three more parameters should be set. First, the number of boosting iterations for the *mboost* algorithm, which is set at 1000 as default. And second, the number of bootstrapping iterations (*bootstrap\_iterations*) performed by the method at each *boosting* function estimation plus the sample size used (*bootstrap\_sampling* - as the fraction of the total number of replicates per scenario) at each bootstrap iteration.

Thus, if one wants to follow the steps used in the manuscript to classify empirical data into the 5 possible outcomes (scenarios), the following commands should be used:

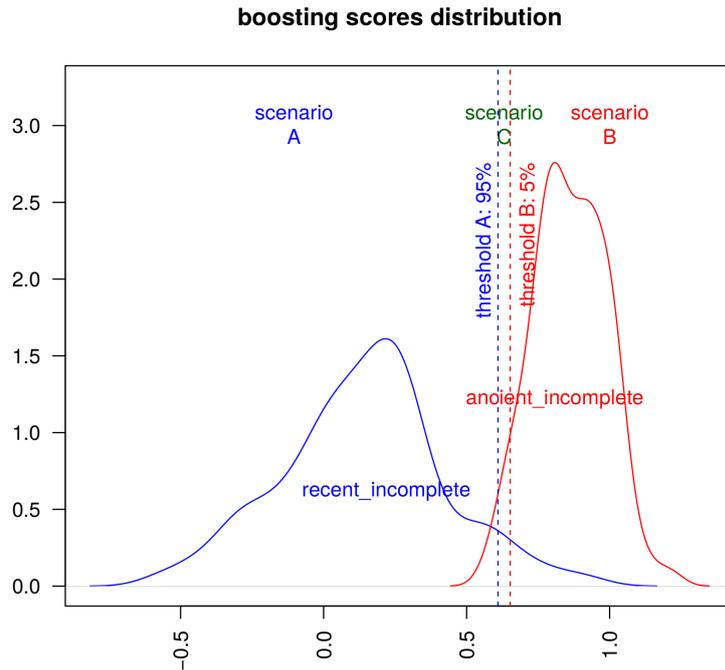
```
> library("hierarchicalBoosting")
```

Load input dataset:

```
> input_path <- system.file("exdata/training_data.txt",
+   package="hierarchicalBoosting")
> input <- read.table(input_path, header=T, stringsAsFactors=F)
```

Define individual *boosting* parameters:

```
> boosting_1 <- c()
> boosting_1["input_data"] <- "original"
```



**Figure 4.** Distribution plot for the *boosting* function #4 (with two thresholds) from the example.

```

> boosting_1["scenario_A"] <- "recent_complete,ancient_complete"
> boosting_1["scenario_B"] <- "neutral,recent_incomplete,ancient_incomplete"
> boosting_1["threshold_scenario_A"] <- FALSE
> boosting_1["threshold_scenario_B"] <- 0.01

> boosting_2 <- c()
> boosting_2["input_data"] <- "boosting_1:scenario_B"
> boosting_2["scenario_A"] <- "neutral"
> boosting_2["scenario_B"] <- "recent_incomplete,ancient_incomplete"
> boosting_2["threshold_scenario_A"] <- 0.99
> boosting_2["threshold_scenario_B"] <- FALSE

> boosting_3 <- c()
> boosting_3["input_data"] <- "boosting_1:scenario_A"
> boosting_3["scenario_A"] <- "recent_complete"
> boosting_3["scenario_B"] <- "ancient_complete"
> boosting_3["threshold_scenario_A"] <- 0.95
> boosting_3["threshold_scenario_B"] <- 0.05

> boosting_4 <- c()
> boosting_4["input_data"] <- "boosting_2:scenario_B"
> boosting_4["scenario_A"] <- "recent_incomplete"
> boosting_4["scenario_B"] <- "ancient_incomplete"
> boosting_4["threshold_scenario_A"] <- 0.95
> boosting_4["threshold_scenario_B"] <- 0.05

```

Create configuration table:

```
> config_table <- data.frame(rbind(boosting_1, boosting_2, boosting_3,  
+ boosting_4), stringsAsFactors=F)
```

Define bootstrapping parameters:

```
> bootstrap_iterations <- 20  
> bootstrap_sampling <- 0.9
```

Check compatibility:

```
> check.HierarchicalBoosting(config_table, input)
```

```
checking scenarios names...ok  
checking significance thresholds...ok  
checking classification tree structure...ok
```

Create *hierarchicalBoosting* object:

```
> hierarchicalBoosting <- train.HierarchicalBoosting(input, config_table,  
+ bootstrap_iterations, bootstrap_sampling)
```

Training Boosting #1

```
recent_complete/ancient_complete vs neutral/recent_incomplete/ancient_incomplete  
input data: original dataset  
number of replicates => scenario A: 199          scenario B: 300  
significance thresholds => scenario A: FALSE      scenario B: 0.01  
bootstrapping.....done
```

Training Boosting #2

```
neutral vs recent_incomplete/ancient_incomplete  
input data: scenario_B from boosting_1  
number of replicates => scenario A: 100          scenario B: 200  
significance thresholds => scenario A: 0.99      scenario B: FALSE  
bootstrapping.....done
```

Training Boosting #3

```
recent_complete vs ancient_complete  
input data: scenario_A from boosting_1  
number of replicates => scenario A: 99           scenario B: 100  
significance thresholds => scenario A: 0.95      scenario B: 0.05  
bootstrapping.....done
```

Training Boosting #4

```
recent_incomplete vs ancient_incomplete  
input data: scenario_B from boosting_2  
number of replicates => scenario A: 100          scenario B: 100  
significance thresholds => scenario A: 0.95      scenario B: 0.05  
bootstrapping.....done
```

Create validation plots:

```
> plot.HierarchicalBoosting(hierarchicalBoosting, config_table, input)
```

Generating validation plots...

```
- boosting_1.plots.pdf
- boosting_2.plots.pdf
- boosting_3.plots.pdf
- boosting_4.plots.pdf
```

Load empirical dataset (selection tests run at SLC45A2 genomic region in CEU):

```
> empirical_path <- system.file("exdata/SLC45A2_CEU_sweep.txt",
+                               package="hierarchicalBoosting")
> empirical <- read.table(empirical_path, header=T, stringsAsFactors=F)
```

Check compatibility:

```
> check.HierarchicalBoosting(config_table, input,
+                             hierarchicalBoosting=hierarchicalBoosting)
```

```
checking scenarios names...ok
checking significance thresholds...ok
checking classification tree structure...ok
checking compatibility: input data and hierarchicalBoosting object...ok
```

Apply *hierarchicalBoosting* object to empirical dataset:

```
> hierarchicalBoosting_results <- apply.HierarchicalBoosting(empirical,
+                                                             config_table, hierarchicalBoosting)
```

Applying hierarchicalBoosting...

```
1 2 3 4
```

Classifying data...

Summarize hierarchicalBoosting results: a signal of 'ancient\_incomplete' selective sweep is detected within the region where the gene is located (SLC45A2: chr5:33,944,721-33,984,780)

```
> classification <- summarize.HierarchicalBoosting(hierarchicalBoosting_results)
```

Summarizing classification...

```
> print(classification)
```

	chromosome	start	end	classification
1	5	33450000	33475000	neutral
2	5	33475000	33500000	neutral
3	5	33500000	33525000	neutral
4	5	33525000	33550000	neutral
5	5	33550000	33575000	neutral
6	5	33575000	33600000	neutral
7	5	33600000	33625000	neutral
8	5	33625000	33650000	neutral

9	5	33650000	33675000	neutral
10	5	33675000	33700000	neutral
11	5	33700000	33725000	neutral
12	5	33725000	33750000	neutral
13	5	33750000	33775000	neutral
14	5	33775000	33800000	neutral
15	5	33800000	33825000	neutral
16	5	33825000	33850000	neutral
17	5	33850000	33875000	neutral
18	5	33875000	33900000	neutral
19	5	33900000	33925000	ancient_incomplete
20	5	33925000	33950000	ancient_incomplete
21	5	33950000	33975000	ancient_incomplete
22	5	33975000	34000000	neutral
23	5	34000000	34025000	neutral
24	5	34025000	34050000	neutral
25	5	34050000	34075000	neutral
26	5	34075000	34100000	neutral
27	5	34100000	34125000	neutral
28	5	34125000	34150000	neutral
29	5	34150000	34175000	neutral
30	5	34175000	34200000	<NA>
31	5	34200000	34225000	<NA>
32	5	34225000	34250000	<NA>
33	5	34250000	34275000	neutral
34	5	34275000	34300000	neutral
35	5	34300000	34325000	neutral
36	5	34325000	34350000	neutral
37	5	34350000	34375000	neutral
38	5	34375000	34400000	neutral

## 4 References

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